ANTITUMOR AGENTS, 157.¹ ABSOLUTE STRUCTURES OF CUMINGIANOSIDES A–F, ANTILEUKEMIC TRITERPENE GLUCOSIDES, AND STRUCTURES OF THE HYDROLYSATES OF CUMINGIANOSIDE A¹

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ABSTRACT.—The stereostructures of cumingianosides A–F, a series of triterpene glucosides with a 14,18-cycloapoeuphane skeleton, have been established by X-ray crystallographic analysis on an aglycone [1C] the acid hydrolysate of cumingianoside A [1], which is a potent cytotoxic triterpene against MOLT-4 human leukemia cells with an EC₅₀ value of <0.00625 μ M. The 14,18-cyclopropane ring in cumingianoside A [1] was opened under acidic conditions in two different directions to give compounds with an apoeuphane skeleton and a dammarane skeleton. Furthermore, it was found that subsequent hydrolysis yielded not only an aglycone with an apoeuphane skeleton [1C] but also an *apo*-rearrangement product [1d].

Our previous investigation of the MeOH extract of the leaves of Dysoxylum cumingianum C. D.C. (Meliaceae) led to the isolation and characterization of cumingianosides A-F [1-6], which are triterpene glucosides with a 14,18-cycloapoeuphane skeleton, along with cumindysosides A [7] and B [8], and trisnortriterpene and tetranortriterpene glucosides as antileukemic principles (2). Cumingianosides A [1] and C [3] demonstrated potent selective cytotoxicity against MOLT-4 human leukemia cells with EC₅₀ values of <0.00625 and <0.0045 μ M, respectively (2). However, the stereochemistry of C-20, C-23, and C-24 in these cumingianosides could not be assigned in the previous study. In order to determine the absolute stereochemistry, X-ray crystallographic analysis was carried out on the hydrolysate of cumingianoside A [1]. This paper describes the determination of the absolute stereostructures of 1-6, as well as the structure of the hydrolysates of 1.

RESULTS AND DISCUSSION

Treatment of deacetylcumingianoside A [1a] with *p*-toluenesulfonic acid in Me₂CO at room temperature yielded deacetylcumingianoside A 23,24-acetonide [1b]. Further reaction of 1b with *p*-toluenesulfonic acid in Me₂CO at reflux furnished two hydrolysates [1c and 1d], together with 1,2,5,6-diisopropylidine-D-glucose [9]. Hydrolysate 1d was acetylated and purified, yielding compound 1e.

¹For Part 156, see Miyahara et al. (1).



The negative fabms of **1c** exhibited an $[M-H]^-$ ion peak at m/z 531, and the molecular formula was confirmed as $C_{33}H_{56}O_5$ by hrfabms. The ¹H-nmr spectrum of **1c** exhibited signals at relatively lowfield due to oxygen-bearing methine groups [δ 3.41 (1H, br s), 3.49 (1H, d, J=8 Hz), 3.89 (1H, br s), and 3.97 (1H, dt, J=2 and 8 Hz)], which were assignable to H-3, H-24, H-7, and H-23, respectively. It also showed, together with a secondary methyl [δ 1.06 (d, J=7 Hz)] and the two methyls of the isopropylidene group [δ 1.39 and 1.40 (each 3H, s)], the presence of seven tertiary methyl groups [δ 0.85, 0.89, 0.95, 1.03, 1.05, 1.17, 1.26 (each 3H, s)]; in contrast, **1** contains only six tertiary methyls. In addition, the absence of cyclopropyl methylene signals, characteristic of 14,18-cycloapoeuphane-type triterpenes, as well as the appearance of **a** olefinic proton signal at δ 5.42 (1H, d, J=2.5 Hz) in the ¹H-nmr spectrum of **1c**, indicated that the cyclopropane moiety in **1b** had opened under the acidic hydrolysis conditions, forming an additional tertiary methyl and a double bond. This conversion is similar to that of a 9,19-cyclolanostane triterpene to its isomeric lanost-9(11)-ene counterpart under acidic conditions (3).

The locations of the additional tertiary methyl group and the double bond were assigned at C-13 and C-14(15), respectively, based on ${}^{1}H^{-1}H$, ${}^{1}H^{-13}C$, and ${}^{1}H^{-13}C$ long-range COSY nmr examination; the ${}^{1}H^{-13}C$ long-range correlations in **1c** are summarized in Figure 1. Furthermore, the absolute stereostructure of **1c** was confirmed unequivocally by X-ray crystallographic analysis (Tables 1 and 2). The data collection information is listed in Table 1. The crystal of **1c** possessed two crystallographically independent molecules, A and B, as shown in Figure 2. Based on the X-ray crystal structure of **1c**, the absolute structure of **1** was determined. Furthermore, the



FIGURE 1. ¹H-¹³C nmr long-range correlations in **1c**.

Chemical formula	C ₃₃ H ₄₆ O ₅		
Formula weight	532.80		
Crystal system	monoclinic		
Space group	$P2_1$		
Z	4		
Cell dimensions			
a (Å)	18.463 (4)		
b (Å)	11.737 (2)		
c (Å)	15.232 (1)		
α (°)	90.00 (1)		
β (°)	108.88 (1)		
γ (°)	90.00 (1)		
$V(Å^3)$	3123.2 (8)		
Crystal size	0.3×0.3×0.3		
d(calcd)(gcm ⁻¹)	1.13045		
F(000)	1172		
$\mu(CuK_a)(mm^{-1})$	0.548		
Max θ (°)	60		
Total reflections	5517		
Unique reflections	4791		
Observed (I>2.30)	4498		
No. of variables	978		
Final R	0.057		
Rw	0.084		

TABLE 1. Crystallographic Data for Compound 1c.

stereostructures of cumingianosides B–F were also established to be as represented by formulas 2-6, respectively, from the chemical correlations of these cumingianosides as shown in Scheme 1 (2).

Compound 1e, the acetate of hydrolysate 1d, gave a $[M]^+$ ion peak at m/z 556 in the fdms, while the negative fabms gave an $[M-CH_3]^-$ ion peak at m/z 541 as the base peak. The elemental composition of this latter peak was analyzed as $C_{34}H_{53}O_5$ by hrfabms, and thus, the molecular formula of 1e was concluded to be $C_{35}H_{56}O_5$. Because 1e possesses an acetyl group as revealed by its ¹H-nmr spectrum [δ 2.07 (3H, s)], the molecular mass of 1d is presumed to be 514 daltons, which is 18 mass units fewer than that of 1c.

The ¹H-nmr spectrum of **1e** exhibited the presence of six tertiary methyl groups [δ 0.76, 0.86, 0.97, 1.11, 1.13, 1.20 (3H each, s)], a secondary methyl group [δ 1.02 (3H, d, J=7 Hz)], two methyls of an isopropyridine group [δ 1.33 and 1.40 (3H each, s)], and an acetyl group [δ 2.07 (3H, s)]. The absence of the cyclopropyl methylene signals suggested that, as in **1c**, the cyclopropane moiety was opened. However, no olefinic proton signal was present in the ¹H-nmr spectrum, suggesting that the cyclopropane moiety had opened in a different manner from that determined for **1c**.

The methine proton signal at $\delta 2.84$ (1H, m) could be assigned to H-20, since this signal was shown to be coupled with the secondary methyl signal at $\delta 1.02$ (d, J=7 Hz), ascribable to Me-20. The chemical shift of H-20 suggested the presence of a double bond at C-17. Therefore, the double bond formed from the opening of the cyclopropane ring moiety was concluded to be at C-13(17).

The ¹H-nmr spectrum of **1e** showed, at relatively lowfield, the existence of three oxygen-bearing methine groups at δ 3.41 (1H, d, J=8.5 Hz), 3.67 (1H, dt, J=2.5 and 8.5 Hz), and 4.67 (1H, br s); the first two of these signals were easily assignable to H-24 and H-23, respectively. From the coupling pattern, the lowest oxygen-bearing methine signal can be assigned to either H-3 or H-7; thus, a methine proton was absent from either C-3 or C-7. The ¹³C-nmr spectrum of **1e** revealed the presence of two tetrasubstituted double bonds [δ 124.8, 131.1, 138.4, and 144.1 (each s)]; one pair must



FIGURE 2. ORTEP view of 1c. (A) molecule A; (B) molecule B.

be due to the double bond at C-13(17). It also exhibited the presence of three oxygenbearing methine carbons [δ 78.3, 76.6, and 75.0] and an oxygen-bearing quaternary carbon (δ 69.6); one less than found in **1c**. These data, together with the difference in the mol wts of 1c and 1e, suggested that the oxygen function at either C-3 or C-7 had been eliminated. Moreover, a three-proton singlet at δ 1.74 in the ¹H-nmr spectrum of 1d indicated the presence of a vinyl methyl group, suggesting that, together with the elimination of the oxygen function, one methyl group had migrated. The locations of the second double bond and the vinyl methyl group were assigned by ¹H-¹³C long-range COSY examination; the ${}^{1}H{}^{-1}C$ long-range correlations in **1e** are shown in Figure 3. The vinyl methyl signal showed ¹H-¹³C long-range correlations with the carbon resonances at δ 124.8 and 138.4. The carbon resonance at δ 138.4 also showed long-range coupling with the methyl signal at δ 1.11, which further exhibited long-range correlations with C-13, C-14, and C-15. Thus, this methyl signal could be assigned as Me-14, with the carbon resonance at δ 138.4 assigned to C-8. Because the vinyl methyl signal also exhibited long-range coupling with the methylene carbon resonance at δ 32.5, assignable to C-6, the location of the double bond is at C-7(8). Therefore, the vinyl methyl was concluded to be at C-7. On the basis of the spectral evidence described above, the structure of 1e was concluded to be as shown in Scheme 2.

Under acidic conditions, the 14,18-cyclopropane ring was opened in two different directions to give compounds with an apoeuphane skeleton (\mathbf{A}) and a dammarane



SCHEME 1

skeleton (**B**). Furthermore, subsequent hydrolysis yielded not only an aglycone with an apoeuphane skeleton (**1c**) but also an *apo*-rearrangement product [**1d**] via the intermediate shown in Scheme 2.

EXPERIMENTAL

GENERAL EXPERIMENTAL PROCEDURES.—All mps were determined on a Fisher-Johns micro meltingpoint apparatus and are uncorrected. Optical rotations were determined with a Rudolph Research Autopol III polarimeter. ¹H- and ¹³C-nmr spectra were obtained on a Bruker AC-300 spectrometer with TMS as internal standard. Fabms measurements were carried out on JEOL JMS-HX110 mass spectrometers.

TREATMENT OF DEACETYLCUMINGIANOSIDE A WITH *P*-TOLUENESULFONIC ACID IN Me_2CO .—A mixture of deacetylcumingianoside A (575 mg), which was prepared in the same way as described in a previous paper (2), and *p*-toluenesulfonic acid (15 mg) in dry Me_2CO (20 ml) was stirred at room temperature for 2 h. The



FIGURE 3. ${}^{1}H^{-13}C$ nmr long-range correlations in **1e**.



product [1b], which crystallized from the reaction mixture, was collected by filtration (colorless needles, 425 mg): mp 205–207°; $\{\alpha\}^{20}$ D –34.2° (c=0.68, MeOH); negative fabms m/z 693 [M–H]⁻; positive-ion fabms m/z 717 [M+Na]⁺; hrfabms m/z calcd for C₃₉H₆₆O₁₀Na 717.4554, found 717.4543; ¹H nmr (pyridine-d₃) δ 0.62, 0.73 (1H each, d, J=5.5 Hz, H-18), 0.93 (3H, s, Me-4 β), 0.95 (3H, s, Me-10), 1.06 (3H, s, Me-8), 1.20 (3H, d, J=6.5 Hz, Me-10), 1.36 (3H, s, Me-4 α), 1.43, 1.52 (3H each, s, Me₂-25), 1.54 (6H, s, isopropylidine-Me₂), 2.59 (1H, d, J=11.5 Hz, H-5), 3.64 (1H, br s, H-3), 3.82 (1H, t, J=7.5 Hz, H-24), 3.99 (1H, m, glucosyl H-5), 4.02 (1H, t, J=8.5 Hz, glucosyl H-2), 4.15 (1H, br s, H-7), 4.18 (1H, t, J=8.5 Hz, glucosyl H-3), 4.26 (1H, t, J=8.5 Hz, glucosyl H-4), 4.35–4.43 (2H, m, H-23 and glucosyl H-6), 4.59 (1H, dd, J=2.5 and 11.5 Hz, glucosyl H-6), 4.91 (1H, d, J=7.5 Hz, glucosyl H-1); ¹³C-nmr data, see Table 3.

HYDROLYSIS OF DEACETYLCUMINGIANOSIDE A 23,24-ACETONIDE BY P-TOLUENE-SULFONIC ACID.—A mixture of 1b (500 mg) and p-toluenesulfonic acid (45 mg) in dry Me₂CO (20 ml) was refluxed overnight with stirring. The reaction mixture was concentrated under reduced pressure to give a residue, which was subjected to Si gel cc. Elution with CHCl₃-MeOH ($60:1 \rightarrow 40:1$) yielded compound **1c** (85 mg), compound 1d(50 mg), and 1,2,5,6-diisopropylidine-D-glucose (45 mg). Compound 1d was further treated with Ac₂O (1 ml) and pyridine (1 ml) at room temperature overnight, and purified by Si gel chromatography [hexane-EtOAc (4:1)] to furnish compound 1e (32 mg) as colorless needles (from hexane). 1c: colorless prisms (from hexane/EtOAc); mp 203–204°; $[\alpha]^{20}D - 118.9^{\circ}$ (c=0.31, CHCl₃); negative-ion fabms 531 (M-H)⁻; positive-ion fabms m/z 555 [M+Na]⁺; hrfabms m/z calcd for C₃₃H₃₆O₅Na 555.4026, found 555.4030; ¹H nmr (CDCl₃) δ 0.85 (3H, s, Me-4 β), 0.89 (3H, s, Me-10), 0.95 (3H, s, Me-4 α), 1.03 (3H, s, Me-14), 1.05 (3H, s, Me-8), 1.06 (3H, d, J=7 Hz, Me-20), 1.17, 1.26 (3H each, s, Me₂-25), 1.39, 1.40 (3H each, s, isopropylidine-Me₂), 2.30 (1H, ddd, J=3.5, 7, and 15 Hz, H-16), 3.41 (1H, br s, H-3), 3.49 (1H, d, J=8 Hz, H-24), 3.89 (1H, br s, H-7), 3.97 (1H, dt, J=2 and 8 Hz, H-23), 5.42 (1H, d, J=2.5 Hz, H-15); ¹³Cnmr data, see Table 3. **1e**: colorless needles (hexane); mp 180–182°; $[\alpha]^{20}$ D – 76.9° (c=0.16, CHCl₂); fdms m/z 556 [M]⁺; positive-ion fabms m/z 541 [M-CH₃]⁺; hrfabms m/z calcd for C₃₄H₅₃O₅ 541.3893, found 541.3891; ¹H nmr (CDCl.) δ 0.76 (3H, s, Me-10), 0.86 (3H, s, Me-4β), 0.97 (3H, s, Me-4α), 1.02 (3H, d, J=7 Hz, Me-20), 1.11 (3H, s, Me-14), 1.13, 1.20 (3H each, s, Me₂-25), 1.33, 1.40 (3H each, s, isopropylidine-Me₂), 1.74 (3H, s, Me-7), 2.06 (3H, s, OAc), 2.24 (1H, dd, J=6.5 and 11.5 Hz, H-16), 2.56 (1H, br d, J=10.5 Hz, H-12), 2.84 (1H, m, H-20), 3.67 (1H, d, J=8.5 Hz, H-24), 3.67 (1H, dt, J=2.5 and 8.5 Hz, H-23), 4.67 (1H, br s, H-3); ¹³C-nmr data, see Table 3.

TABLE 2. Fractional Atomic Coordinates ($\times 10^4$) and Equivalent Isotropic Temperature Factors for Nonhydrogen Atoms of Compound **1c** (**A** and **B** forms) with Their Estimated Standard Deviations in Parentheses.

Atom	x	у	Z	Beq (Å ²)
O-3A	8878 (2)	5960 (4)	7586 (3)	6.14 (13)
O-7A	6202 (2)	5530 (3)	5525 (3)	4.23 (9)
O-23A	1906 (2)	8862 (3)	6815 (4)	5.74 (13)
O-25A	1706 (2)	5671 (4)	7450 (3)	5 33 (12)
C-1A	7898 (3)	8115 (5)	7386 (4)	4.90 (15)
C-2A	8748 (3)	8007 (6)	7464 (5)	5.53 (17)
C-3A	8919 (3)	6940 (5) 6703 (5)	7022 (4)	4.94 (16)
C-5A	7572 (3)	6966 (4)	5948 (3)	3.64 (12)
C-6A	6997 (3)	6737 (5)	4977 (4)	4.22 (13)
C-7A	6198 (3)	6568 (4)	5015 (3)	3.59 (12)
C-7A	5909 (2) 6522 (2)	7391 (4) 7894 (4)	5450 (3) 6398 (3)	3.10 (10)
C-10A	7360 (3)	8067 (4)	6389 (4)	3.82 (12)
C-11A	6249 (3)	8838 (5)	6916 (4)	4.53 (15)
C-12A	5384 (3)	8898 (4)	6741 (4)	3.94 (13)
C-14A	5170 (2)	7298 (4)	5618 (3)	2.96 (10)
C-15A	4575 (3)	6741 (4)	5049 (3)	3.61 (12)
C-16A	3879 (3)	6815 (4)	5358 (3)	3.67 (12)
C-1/A	4080 (2)	(4) (4) (5)	7265 (3)	3.22 (12)
C-19A	7454 (3)	9159 (5)	5879 (5)	5.01 (17)
C-20A	3593 (2)	7918 (4)	6675 (3)	3.38 (11)
C-21A	3764 (3)	8990 (5)	7274 (4)	4.88 (16)
C-23A	2198 (2)	7762 (4)	6690 (3)	3.85 (12)
C-24A	1484 (3)	7079 (4)	6228 (3)	3.84 (12)
C-25A	1514 (3)	5816 (4)	6465 (4)	4.27 (14)
C-26A	/46 (4) 2165 (4)	52/0 (6)	5945 (6) 6202 (5)	6.96 (23)
C-28A	8518 (3)	5557 (6)	5688 (6)	6.28 (21)
C-29A	8708 (3)	7596 (6)	5392 (5)	6.09 (20)
C-30A	5727 (3)	8586 (5)	4743 (4)	4.44 (14)
C-32A	1229 (4)	8709(5)	7925 (5)	4.55 (14)
C-33A	657 (4)	9597 (6)	6348 (5)	6.13 (20)
O-3B	7223 (3)	5055 (4)	7464 (3)	6.64 (14)
0-/B	5/69(2) 709(2)	4925 (4)	9649 (4) 7835 (3)	6.01 (13) 5 22 (11)
O-24B	-59 (2)	3092 (4)	8414 (3)	5.47 (11)
O-25B	473 (2)	5295 (3)	8233 (3)	5.14 (11)
C-1B	6378 (3) 7150 (3)	2818 (5)	7444 (4) 7205 (á)	4.50 (14)
C-3B	7591 (3)	3989 (5)	7846 (4)	4.90 (16)
C-4B	7672 (3)	3972 (4)	8882 (3)	3.91 (13)
C-5B	6869 (3)	3730 (4)	8982 (3)	3.44 (11)
С-ов	6062 (3)	3854 (6)	9985 (4) 10030 (4)	4.62 (14)
C-8B	5546 (2)	2862 (4)	9519 (3)	3.60 (11)
C-9B	5597 (2)	2761 (4)	8527 (3)	3.39 (11)
С-10В	6425 (2) 5033 (3)	2681 (4) 1884 (5)	8462 (3) 7922 (4)	3.47 (11) 4 79 (15)
C-12B	4270 (3)	1803 (5)	8092 (4)	4.46 (14)
C-13B	4053 (2)	2841 (4)	8562 (3)	3.33 (11)
C-14B	4706 (3) 4442 (3)	3087 (4) 3/30 (6)	9438 (3)	3.74 (12)
C-16B	3590 (3)	3385 (6)	9826 (4)	5.28(17)
C-17B	3380 (3)	2619 (4)	8960 (3)	3.88 (12)
C-18B	3884 (3)	3890 (5)	7922 (4)	4.36 (14)
C-19B	0812 (3) 2551 (2)	1520 (4)	8820(5) 8308(3)	4.96 (16)
C-21B	2344 (3)	2031 (6)	7452 (4)	5.53 (17)
C-22B	1983 (3)	2568 (5)	8836 (4)	4.30 (13)
C-25B C-24B	1168 (3)	2980 (4) 3406 (5)	8311 (4) 8012 (4)	3.92 (12)
C-25B	740 (3)	4689 (5)	9096 (4)	4.63 (15)
C-26B	251 (4)	4996 (7)	9706 (5)	6.70 (23)
C-27B	1574 (4)	5069 (6)	9556 (5)	6.51 (21)
C-29B	8296 (3)	3079 (6)	9363 (4)	5.17 (16)
C-30B	5773 (3)	1762 (6)	10115 (5)	6.01 (19)
C-31B	-63 (3)	2275 (5)	7731 (4)	4.63 (15)
C-33B	-431(4) -430(5)	2/90 (8) 1187 (6)	0/81() 7921(7)	7.76 (25)

Carbon	Compound				
	1'	1 b ^b	1c*	1e'	
1	34.2	33.9	32.6	32.4	
2	22.7	26.4	25.0	23.0	
3	76.0	75.5	76.2	78.3	
4	36.4	37.8	37.0	36.2	
5	41.5	40.3	40.5	44.5	
6	20.4	21.0	23.7	32.5	
7	78.6	78.4	72.4	124.8	
8	34.8	35.4	44.4	138.4	
9	45.0	45.3	41.8	52.9	
10	37.3	37.9	37.6	35.3	
11	17.0	17.4	16.4	25.6	
12	27.4	28.0	33.7	23.6	
13	26.8	27.2	47.0	144.1	
14	38.8	39.5	162.5	52.5	
15	24.1	25.2	119.4	27.7	
16	25.5	26.4	35.3	40.5	
17	52.8	53.1	60.8	131.1	
18	17.4	17.4	18.5	24.6	
19	16.0	16.5	15.2	14.1	
20	31.9	35.3	33.8	28.9	
21	19.4	20.4	20.1	20.3	
22	36.9	39.9	41.7	40.2	
23	69.9	78.8	77.2	75.0	
24	75.1	88.9	87.7	86.6	
25	74.2	69.8	69.8	69.6	
26	26.1	26.9	24.7	24.7	
27	27.3	27.6	27.9	27.7	
28	27.9	29.0	28.2	27.4	
29	21.8	22.9	22.1	21.6	
30	20.0	20.5	27.8	22.0	
Isopropylidine					
CH,		27.6, 28.0	27.6, 27.2	27.7, 26.8	
c		108.5	108.5	107.6	
Glucosvl					
1'	98.6	100.7			
2'	74.0	75.5			
3'	78.6	78.7			
4'	70.2	72.5			
5'	73.3	78.1			
6'	63.3	63.5			
Ac	20.8, 21.3			21.3, 170.8	
	170.9, 171.7			-,	

TABLE 3. ¹³C-Nmr Data (δ) for Compounds 1, 1b, 1c, and 1e (75.5 MHz).

^{*}Measured in CDCl₃.

^bMeasured in pyridine-d₅.

X-RAY CRYSTALLOGRAPHY.²—X-ray diffraction data were measured on an Enraf-Nonius CAD4 diffractometer. Intensity data collection was accomplished by the ω -2 θ scan method with graphite-monochromated CuK α radiation up to θ =60°; 4498 unique reflections with $I>2.3\sigma$ were used for refinement. The structure was solved by the direct methods program SIR88.

²Hydrogen coordinates, thermal parameters, bond distances and angles, and observed and calculated structure factors have been deposited with the Cambridge Crystallographic Data Centre and can be obtained upon request from Dr. Olga Kennard, University Chemical Laboratory, 12 Union Road, Cambridge CB2 1EZ, UK.

All atomic parameters, with anisotropic temperature factors for non-hydrogen atoms and isotropic ones for hydrogen atoms, were refined by a block-diagonal least-squares method. The final *R* value was 0.057. Crystallographic data are given in Table 1, while final fractional atomic coordinates are shown in Table 2.

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